

Potential Natural Antibacterial Agent for *P. gingivalis* Periodontitis Infection: A Comprehensive Review of Source, Structure and Mechanism actions

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ABSTRACT

Background: the pathogenic bacteria *P. gingivalis* grows in the oral cavity. This bacterium could attack immune system which lead to inflammation of most tissues. *P. gingivalis* can cause a variety of serious and dangerous condition such as periodontitis, Alzheimer, rheumatoid arthritis, diabetes, and pneumonia. Antibiotics have been used for years as a treatment against this bacterium, like metronidazole, amoxicillin, and clindamycin.

Method: *P. gingivalis* is reported to be resistant to these antibiotics, thus exploration to discover alternatives has been demanded.

Result: natural product compounds are known to have antibacterial activity and cause fewer side effects. Turmeric, eucalyptus, and several other plants have been reported to have antibacterial activity against *P. gingivalis* with a MIC of 1g/mL from an ethyl acetate leaf extract of eucalyptus.

Conclusion: Decent antibacterial activity could be used as a reference to discover new drugs as alternatives against *P. gingivalis*.

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INTRODUCTION

Dental disease can become a chronic disease if not treated properly. Chronic diseases of the teeth and dentition are associated with gingivitis or periodontitis which can cause inflammation and damage the bone supporting the teeth. The inflammation can lead to complications and form periodontal pockets between the gums and teeth.¹ Periodontal disease is also linked to other diseases such as cardiovascular disease and diabetes.² This dental infection is caused by various types of bacteria such as *Porphyromonas Gingivalis*, *Aggregatibacter actinomycetemcomitans*, and *Fusobacterium nucleatum*.³ However, *P. gingivalis* is a known as pathogenic bacteria that plays a major role in periodontal disease.^{4, 5} *P. gingivalis* infects teeth through the action of the cysteine proteases lysin and arginine-gingipain during the initiation and progression of inflammation.⁶ Studies on the effect of *P. gingivalis* on periodontal disease have been conducted on mice as test animals that showed gingival inflammation and alveolar bone loss.⁷

P. gingivalis is a Gram-negative bacterium which incorporated with “red complex”. This group of bacteria consist of *P. gingivalis*, *Treponema denticola*, and *Tannerella forsythia*.^{8, 9} *P. gingivalis* has been reported as the cause of periodontitis or tooth loss disease.^{10, 11} In 2020 Mei et al and Olsen et al reported that *P. gingivalis* has a role in several dangerous diseases such as Alzheimer’s, rheumatoid arthritis, diabetes, and pneumonia.^{12, 13} The high complexity of the diseases caused by *P. gingivalis* showed the urgency of discovering alternative treatments.

P. gingivalis has shown significant resistance against commercial antibiotics such as 21.56% against metronidazole, 25.49% against amoxicillin, and 23.52% against clindamycin.¹⁴ Thus, antibacterial exploration has become important. Curcumin, a phenolic compound found in turmeric, has been shown to exhibit antibacterial activity against *P. gingivalis*, which causes periodontitis inflammation.¹⁵ Aside from curcumin, phloroglucinol-sesquiterpene-coupled, which is isolated from eucalyptus, showed antibacterial activity to inhibit the growth of *P. gingivalis*.¹⁶ The emphasis of this review is on natural compounds having antibacterial activity against *P. gingivalis*. In this review, we also discuss the isolation technique and bioassay of natural compounds against *P. gingivalis*. Furthermore, we also discussed the treatment mechanism produced by *P. gingivalis*.

***P. gingivalis* BACTERIA**

P. gingivalis is an anaerobe Gram-negative bacterium which is attached to liposaccharide-lipid A at its membrane. This lipid formed a thin layer which stimulated the inflammation on the cell.¹⁷ Lipopolysaccharide in *P. gingivalis* is hard to dissociate, so that causes drug resistance.¹⁸ The growth of this bacteria in the cell is supported by arginine amino acid and lysingipains produced by its reproduction protein. Proliferation will be prolonged if Arg and Lys gingipains are inhibited. When *P. gingivalis* attacks human albumin serum, it becomes *P. gingivalis* proliferate media. As a result, *P. gingivalis* caused a disease complications since albumin is producing blood plasma protein.¹⁹ *P. gingivalis* will inflame every area of the human body since it is transmitted by the blood. *P. gingivalis* has been linked to some diseases such as pneumonia, pancreatic cancer, gastrointestinal, periodontitis or tooth loss.

TREATMENT MECHANISM OF DISEASE CAUSED BY *P. gingivalis*

2.1 Periodontitis

Periodontitis, or tooth loss, is mostly caused by the bacterium *P. gingivalis*. This bacterial growth inside the oral cavity uses albumin in human blood as a media.^{12, 19} More than 200 million adults all around the world have been attacked by these bacteria and have suffered great inflammation, which leads to periodontitis.²⁰ Periodontitis mechanisms start with *P. gingivalis*, which has been growing on the oral cavity surface exploiting complement growth media (red and white molecules). The rapid growth rate caused other oral bacteria to change their composition and quantity so that brown shafts appeared. Significant composition changes cause inflammation periodontitis, which is characterized by an exfoliated oral cavity. Chronic periodontitis will cause tooth loss.⁹ Periodontitis which caused by *P. gingivalis* is called a virulence factor. Virulence factor is a damaged host which is caused by molecules or organism (bacteria, fungi, virus, and protozoa).²¹ Antibiotics that have been used as a treatment for periodontitis are amoxicillin, metronidazole, azithromycin, and moxifloxacin. However, these antibiotics are reported to be resistant against *P. gingivalis*.²² Therapeutic method has also been developed like Cortexyme Inc® corporation does as a periodontitis preventive measure.²³

2.2 Alzheimer

Alzheimer's disease is a neurodegenerative disease that causes memory and thinking ability loss. One of the causes of this disease is the expression of a toxic active proteolytic protease protein named gingipains from *P. gingivalis*. Gingipains caused amyloid-protein (APP) precursor or amyloid shrieved and neurofibrillary cleavage, resulting in normal nerve system degeneration.^{24, 25} Nerve cell death which caused by gingipains could be induced by a small molecule such as COR286 and COR271. Commercial antibiotics such as moxifloxacin and doxycycline have been tested *in vitro*, yet they are still not effective enough to protect nerve cells from gingipains.²⁶

2.3 Rheumatoid Arthritis

Rheumatoid arthritis (RA), or joint inflammation, is an inflammation chronic disease. The late treatment could lead to paralysis. This disease is carried by the presence of autoimmune induced bacteria *P. gingivalis*.²⁷ RA mechanism starts with the peptidyl arginine deiminase (PAD) enzyme produced by *P. gingivalis*. The PAD enzyme is an enzyme that produces citrullinated protein, which is known to be the cause of RA. Thus, this mechanism determines the role of *P. gingivalis* in inducing RA through PAD production.^{28, 29}

Chronic RA treatments have been developed to lessen paralysis and disability. DMARD (Disease Modifying Antirheumatic Drug) therapeutic already known as an effective initial treatment for RA. The combination of DMARD with conventional drugs such as methotrexate, glucocorticoids is reported to be more effective than DMARD alone. However, commercial drugs could provoke another disease. Exploration of natural compounds is necessary to increase the effectiveness treatment of RA along with DMARD.^{30, 31}

2.4 Diabetes

Diabetes has been a familiar deathly disease to us. Aside from genetic, the disease could be caused by bad dietary. High sugar blood level is a potential environment for bacteria to growth such as *P. gingivalis*.³² *In vivo* study on rat male adult reported that adding *P. gingivalis* could rise the sugar blood level.³³ The main mechanism of how *P. gingivalis* could rise the sugar blood level still remain unknown. Deteriorate of periodontitis and diabetes due to *P. gingivalis* requires treatment to each disease since we cannot treat them at once before knowing the mechanism.³⁴

2.5 Pneumonia

Pneumonia is a deathly inflammation disease. The inflammation of this disease is due to induction of anaerobe bacteria such as *P. gingivalis*. *P. gingivalis* has the ability to manipulate the immune system of host through gingipains.¹⁰ Although anaerobe bacteria cannot survive in warm temperature such as the lungs, gingipains still able to induce interleukin (IL)-8 and IL-6. IL-6 and IL-8 are cytokine secretion of inflamed the lungs. By the induction of gingipains on IL-6 and IL-8, *P. gingivalis* takes on the role of pneumonia cause.³⁵

The treatment can be achieved through preodontal therapy. The oral cavity, being the initial site for the growth of *P. gingivalis* is treated to suppress its proliferation. This therapy could reduce the number of *P. gingivalis* thus suppress the effect on pneumonia.³⁶

NATURAL PRODUCTS AS ANTIBACTERIAL AGENT OF *P. gingivalis*

3.1 Phenolic

Phenolic compounds are generally known to have many hydroxyl groups. In the mechanism of inhibiting pathogenic bacteria, hydroxyl groups play an important role in damaging the bacterial cell membrane. Especially Gram-negative bacteria have structures that are coated with an outer membrane and cover the cell wall externally. The outer membrane can block antibiotics from entering the cell because it is composed of lipopolysaccharides. Hydroxyl groups on phenolic compounds can affect the integrity of the cell membrane by releasing protons.³⁷ In addition, there are porin channels in the bacterial cell membrane that can facilitate hydrophilic antibacterials to diffuse into the cell.³⁸ Phenolic compounds can also inhibit bacterial cells through this system.³⁹ The activity of phenolic compounds against *P. gingivalis* are described in Table 1. The structure of these compounds is shown in Figure 1.

Table 1. Activity of phenolic compounds from plant materials against *P. gingivalis*

Plants	Extract	MIC ($\mu\text{g/mL}$)	Reference
Turmeric (<i>Curcuma longa</i>)	Curcumin (1)	62.5	40
		12.5	41, 42
		125	39
		20 (suppress percentage 40%)	43, 44
<i>Cinnamomum zeylanicum</i> essential oil	Cinnamaldehyde (2)	2 μM	45
<i>Monechma ciliatum</i> seed	Coumarin (3)	50	46
<i>Zingiber officinale</i> Roscoe	10-gingerol (4)	6	47
	12-gingerol (5)	15	47
<i>Amphipterygium adstringens</i> bark	6-(16'-Z-nonadecenyl)-salicylic acid (6) 6-(8'-Z-pentadecenyl) salicylic acid (7) 6-nonadecenyl salicylic acid (8) 6-pentadecyl salicylic acid (9)	12	48
		18	
		70	
		126	
<i>Olea europaea</i>	Oleuropein (10) Hydroxytyrosol (11) Oleocanthal (12) Oleacin (13)	625 $\mu\text{M/mL}$	49, 50
		156 $\mu\text{M/mL}$	
		156 $\mu\text{M/mL}$	
		312 $\mu\text{M/mL}$	
<i>Citrus limon</i> peel	8-geranyloxypsolaren (14) 5-geranyloxypsolaren (15) 5-geranyloxy-7-methoxycoumarin (16)	0.3 μM	47
		0.15 μM	
		0.45 μM	
<i>Magnolia officinalis</i>	Honokiol (17) Magnolol (18)	25 $\mu\text{M/mL}$	51
		25 $\mu\text{M/mL}$	
Grapes skin	Resveratrol (19)	78.12-156.25 $\mu\text{M/mL}$	52
<i>Acronychia baueri</i> Schott	3-(4'-geranyloxy-3'-methoxyphenyl)-2-trans-propenoic acid (20)	31.3 as antibiofilm	53
<i>M. paniculate</i>	Murrangatin (21) Murrangatin acetate (22) Micropubescin (23)	100	54
		100	
		100	
Licorice root	Licochalcone A (24)	10	55

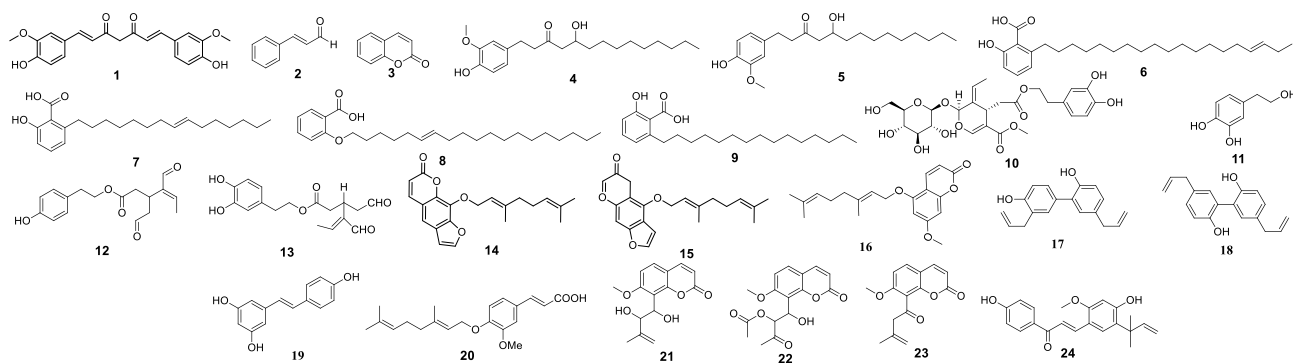


Figure 1. Phenolic Structure of curcumin (1), Cinnamaldehyde (2), Coumarin (3), 10-gingerol (4), 12-gingerol (5), 6-(16Z-nonadecenyl)-salicylic acid (6), 6-(8Z-pentadecenyl) salicylic acid (7), 6-pentadecyl salicylic acid (8), oleuropein (10), hydroxytyrosol (11), oleocanthal (12), oleacin (13), 8-geranyloxypsolaren (14), 5-geranyloxypsolaren (15), 5-geranyloxy-7-methoxycoumarin (16), Honokiol (17), magnolol (18), resveratrol (19), 3-(4'-geranyloxy-3'-methoxyphenyl)-2-*trans*-propenoic acid (20), Murrangatin (21), murrangatin acetate (22), micropubescin (23), licochalcone A (24)

3.2 Terpenoid

In terpenoid compounds, the mechanisms studied for bacterial inhibition can be through two processes: oxygen uptake and oxidative phosphorylation. In aerobic microbes, growth is affected by the concentration of available oxygen. As for bacterial cell respiration in the cytoplasmic membrane, a biochemical process called oxidative phosphorylation occurs.⁵⁶ The presence of terpenoid compounds can have chemical interactions on the cytoplasmic membrane that cause the release of oxidative phosphorylation thus changing the cellular respiration of bacteria.⁵⁷ In another study it was reported that terpenoids can provide bacteriostatic effects through carbonylation, but cannot provide bactericidal effects. In addition, lipophilicity, water solubility, and the presence of hydroxyl groups in the terpenoid structure also affect the antibacterial action so that terpenoids have potential as antiseptics.⁵⁸ Some mechanisms proposed for diterpenes like hardwickiic was the compound may block cell membranes due to their lipophilicity structure. The lipophilicity aids insertion into the cell membrane, and the presence of a hydrophilic group that interacts with a phosphorylated group on the membrane may increase bacterial lysis.⁵⁹ The activity of terpenoid compounds against *P. gingivalis* are described in Table 2. The structure of these compounds is shown in Figure 2.

Table 2. Activity of terpenoid compounds from plant materials against *P. gingivalis*

Plants	Extract	MIC ($\mu\text{g/mL}$)	Ref.	
<i>Eucalyptus globulus</i>	Macrocarpal A (25)	0.39	60	
	Macrocarpal B (26)	0.78		
	Macrocarpal C (27)	0.2		
	Macrocarpal D (28)	0.39		
	Macrocarpal H (29)	0.78		
	Macrocarpal I (30)	6.25		
	Macrocarpal J (31)	6.25		
	Eucalyptone (32)	1.56		
	<i>Illicium lanceolatum</i>	α -santal-1en-10-one (33)		5-20
<i>P. linteus</i> fruit		<i>trans</i> - γ -monocyclofarnesol (34)	5.9	62
		γ -ionylideneacetic acid (35)	34.1	
<i>Copaifera</i> spp	Polyalthic acid (36)	3.12-12.5	63	
	Hardwickiic acid (37)	6.25		
	Kaurenoic acid (38)	1.59-12.5		
	<i>ent</i> -hardwickiic acid (39)	>10		
	13 <i>E-ent</i> -labda-7,13-dien-15-oic-acid (40)	>10		

<i>Mikania glomerata</i>	Kaurenoic acid (38)	6.25-12.5	64
<i>Vitis vinifera</i>	Maslinic acid (41)	4.9	65
	24Z-isomasticadienolic acid (42)	2.4	
	Oleanolic acid (43)	9.9	
	Oleanonic aldehyde (44)	625	
	Copalic acid (45)	3.1	66
<i>Copaifera langsdorfii</i>	Ceanothic acid (46)	62	67
	27-hydroxyceanothetric acid (47)	1250	
	Phloroglucinol-sesquiterpene (48)	11.5 mm (inhibition zone)	68
<i>Myrmecodia pendans</i>	R1-barrigenol (49)	100 (MBC)	69
<i>P. tobira</i>	Xanthorrhizol (50)	6.8	70
<i>Iostephane heterophylla</i>	<i>ent</i> -pimara-8(14),15-dien-19-oic acid (51)	1.25	71
	<i>ent</i> -8(14),15-pimaradien-3 β -ol (52)	10	
	Sodium salt of compound 51 (53)	1	
	Deacetyl laurenobiolide (54)	500	72
<i>Laurus nobilis</i> L	Laurenobiolide (55)	250	
	12-ethoxynimbolinins C (56)	15.6	73
<i>Melia toosendan</i>	1-cinnamoyltrichilin (57)	31.3	

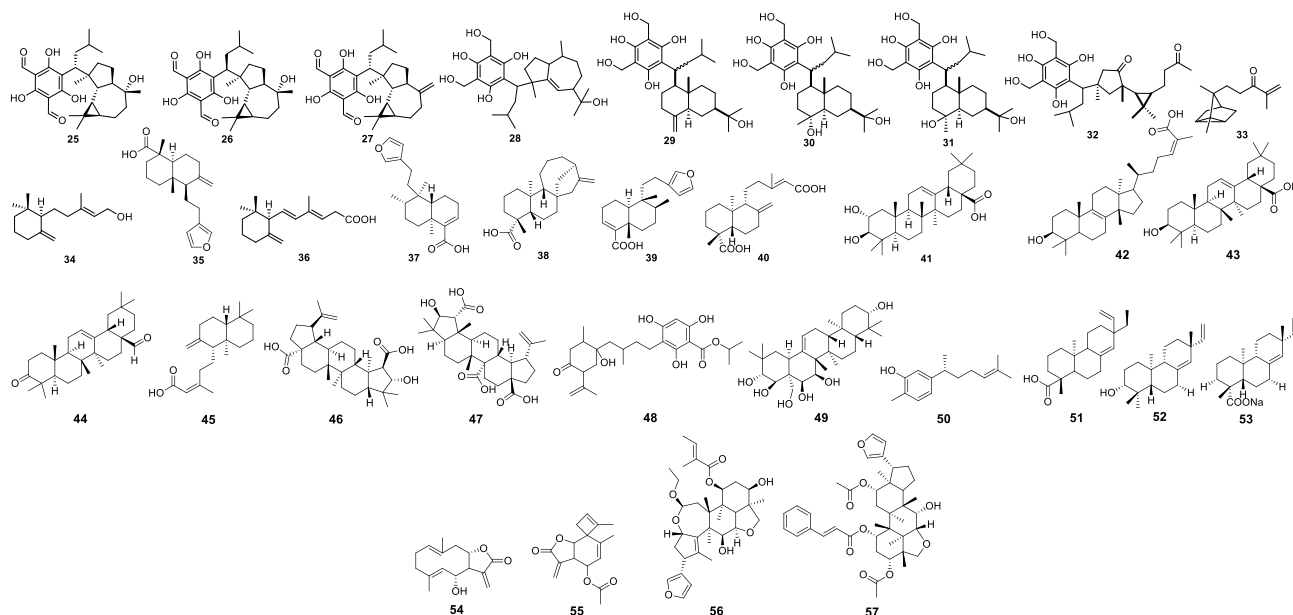


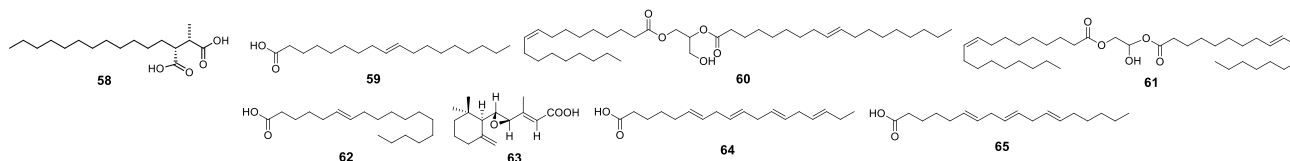
Figure 2. Terpenoid Structure of macrocarpals A (**25**), macrocarpals B (**26**), macrocarpals C (**27**), macrocarpals D (**28**), macrocarpals H (**29**), macrocarpals I (**30**), macrocarpals J (**31**), eucalyptone (**32**), α -santal-1en-10-one (**33**), *trans*- γ -monocyclofarnesol (**34**), γ -ionylideneacetic acid (**35**), Polyalthic acid (**36**), hardwickiic acid (**37**), kaurenoic acid (**38**), *ent*-hardwickiic acid (**39**), 13*E-ent*-labda-7,13-dien-15-oic-acid (**40**), Maslinic acid (**41**), 24Z-isomasticadienolic acid (**42**), oleanolic acid (**43**), oleanonic aldehyde (**44**), copalic acid (**45**), ceanothic acid (**46**), ceanothetric acid (**47**), phloroglucinol-sesquiterpene (**48**), R1-barrigenol (**49**), Xanthorrhizol (**50**), *ent*-pimara-8(14),15-dien-19-oic acid (**51**), *ent*-8(14),15-pimaradien-3 β -ol (**52**), sodium salt of 54 (**53**), deacetyl laurenobiolide (**54**), laurenobiolide (**55**), 12-ethoxynimbolinins C (**56**), 1-cinnamoyltrichilin (**57**).

3.3 Fatty Acid

Fatty acids can damage bacterial cells through several mechanisms: detergent activity that causes membrane disruption, plasma membrane interaction with fatty acids, transport of fatty acids into the cytosol through the cell membrane, and chemical interaction of proteins in the cell membrane with fatty acids. These mechanisms can cause cell lysis, formation of pores in cells, disruption of protein binding, and changes in cell membranes.^{74, 75} The activity of fatty acids against *P. gingivalis* are described in Table 3. The structure of these compounds is shown in Figure 3.

Table 3. Activity of fatty acids from plant materials against *P. gingivalis*

Plants	Extract	MIC ($\mu\text{g/mL}$)	Ref.
<i>Roccella fuciformis</i>	(+)-roccellic acid (58)	46.9	76
<i>Monechma ciliatum</i> seeds	Oleic acid (59)	≤ 15	46
	1,2-dioleoylglycerol (60)	≤ 100	
	1,3-dioleoylglycerol (61)	100	
<i>Foeniculum vulgare</i>	Petroselinic acid (62)	5	77
<i>P. linteus</i>	Phellidene fatty acid E (63)	155	62
<i>Enteromorpha linza</i>	Stearidonic acid (64)	9.76	78
	γ -linolenic acid (65)	9.76	

**Figure 2.** Fatty Acid roccellic acid (**58**), oleic acid (**59**), 1,2-dioleoylglycerol (**60**), 1,3-dioleoylglycerol (**61**), petroselinic acid (**62**), phellidene fatty acid E (**63**), stearidonic acid (**64**), γ -linolenic acid (**65**)

3.4 Flavonoid

Flavonoids contain prenyl groups, alkyl chains, alkylamino, and heterocyclic groups that affect different antibacterial mechanisms depending on each group in the structure. The main mechanism as an antibacterial is through interaction with the cell membrane. The lipophilicity or hydrophilicity of the flavonoid structure determines the interaction that occurs on the outside or inside of the cell bilayer.⁷⁹ The more polar flavonoid compounds will penetrate the bacterial cell wall, then break down cell proteins and damage the cytoplasmic membrane, it leads the bacterial cell death.⁸⁰ The activity of flavonoid compounds against *P. gingivalis* are described in Table 4. The structure of these compounds is shown in Figure 4.

Table 4. Activity of flavonoids from plant materials against *P. gingivalis*

Plants	Extract	MIC ($\mu\text{g/mL}$)	Ref.
Rhubarb root	Rhein (66)	2500	81
Green tea	Epigallocatechin (67)	125-500	82
<i>Pelargonium sidoides</i>	Proanthocyanidins (68)	90%	83
<i>Kaempferia pandurata</i> root	Panduratin A (69)	4	84
<i>Glycyrrhiza uralensis</i>	Licorisoflavan A (70)	1.56	85
	Licorisoflavan C (71)	6.25	
	Licorisoflavan D (72)	6.25	
	Licorisoflavan E (73)	12.5	
	Licoricidin (74)	3.125	
	Biflorin (75)	625	
	Kaempferol (76)	20	
<i>Syzygium aromaticum</i>	Rhamnocitrin (77)	625	86
	Myricetin (78)	20	
	Kaempferol (74)	25-100	
	epicatechin-(4 β →8)-epicatechin-(4 β →8,2 β →O→7)-catechin (79)	100	
<i>Polygonum tinctorium</i>	Kaempferol (74)	25-100	87
<i>Vaccinium vitis-idaea</i> L.	epicatechin-(4 β →8)-epicatechin-(4 β →8,2 β →O→7)-catechin (79)	100	88
Perilla seed	Luteolin (80)	12.5-50	89
<i>M. flabellifolia</i>	3',4',5',7-tetramethoxyflavone (81)	100	90
	Flavan-3-ol (82)	100	
<i>Lonicera caerulea</i>	Cyanidin-3-O-glukosida (83)	36.3% for biofilms	91

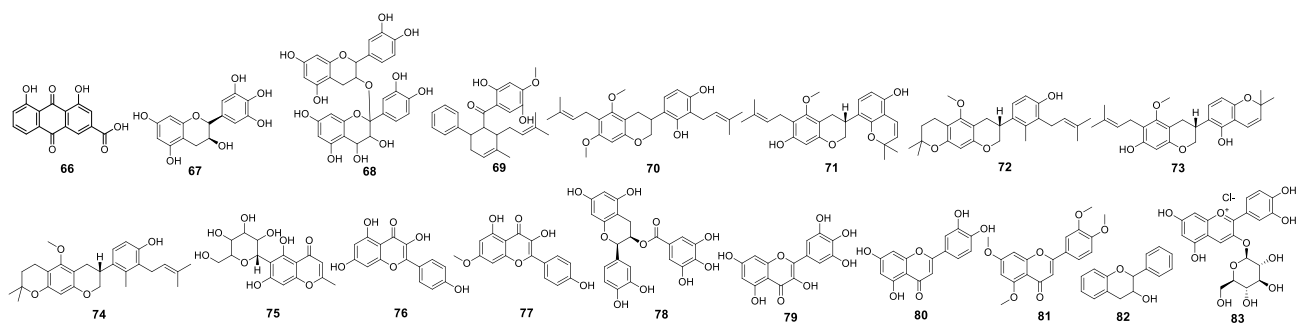


Figure 4. Flavonoid structure of rhein (**66**), epigallocatechin (**67**), Proanthocyanidins (**68**), panduratin A (**69**), licorisoflavan A (**70**), licorisoflavan C (**71**), licorisoflavan D (**72**), licorisoflavan E (**73**) and licoricidin (**74**), Biflorin (**75**), kaempferol (**76**), rhamnocitrin (**77**) and myricetin (**78**), epicatechin-(4 β →8)-epicatechin-(4 β →8,2 β →O→7)-catechin (**79**), Luteolin (**80**), 3',4',5',7-tetramethoxyflavone (**81**), flavan-3-ol (**82**), cyanidin-3-O-glukosida (**83**).

3.5 Other Compounds

In general, there are many other secondary metabolite compounds in plants. One of them is alkaloid compounds which also have potential as antibacterials. In bacterial cell walls, alkaloids can prevent the formation of cell layers, so that they do not form completely and cause death.⁸⁰ In addition, based on the biosynthetic pathway, alkaloids are formed from amino acids, so they can form chemical interactions with DNA in bacteria that cause cell division disorders. This can also lead to bacterial cell death.⁹² The activity of alkaloids and the other compounds against *P. gingivalis* are described in Table 5. The structure of these compounds is shown in Figure 5.

Table 5. Activity of other compounds from plant materials against *P. gingivalis*

Plants	Extract	MIC ($\mu\text{g/mL}$)	Ref.
<i>Var. druidarum</i>	Hypoprotocetraric acid (84)	250	76
<i>I. tinctoria</i>	Tryptanthrin (85)	6.25-12.5	93
<i>Vitis vinifera</i>	5-hydroxymethyl-2-furfural (86)	16	65
<i>Solenostemma argel</i> leave	Argeloside I (87)	60	94
<i>Garcinia kola</i> seed	8E-4-geranyl-3,5-dihydroxybenzophenone (88)	62.5	95
	δ -garcinoic acid (89)	31.3	
<i>M. toosendan</i>	Trichilin B (90)	31.5	73
<i>Piper cubeba</i>	(-)-Cubebin (91)	400	96
	(-)-O-methylcubebin (92)	200	
	(-)-O-benzylcubebin (93)	300	
	(-)-O-acetylcubebin (94)	>400	
<i>M. toosendan</i> fruit	1 α -triglyoxy-3 α -acetoxyl-7 α -hydroxyl-12 β -ethoxynimbolinin (95)	31.25	97

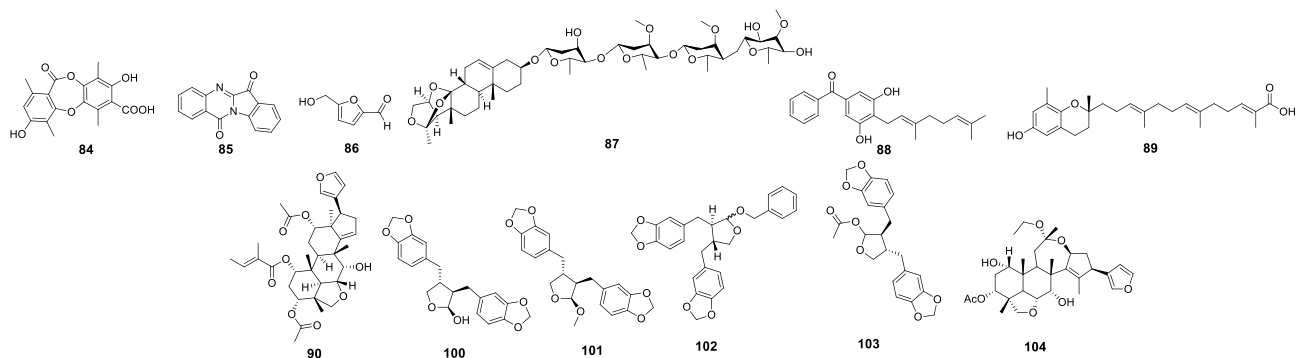


Figure 9. Other Compounds Structure hypoprotocetraric acid (**84**), Tryptanthrin (**85**), 5-hydroxymethyl-2-furfural (**86**), argeloside I (**87**), 8E-4-geranyl-3,5-dihydroxybenzophenone (**88**), δ -garcinoic acid (**89**), Trichilin B (**90**), (-)-Cubebin (**100**), (-)-O-methylcubebin (**92**), (-)-O-benzylcubebin (**93**), (-)-O-acetylcubebin (**94**), and 1 α -triglyoxy-3 α -acetoxyl-7 α -hydroxyl-12 β -ethoxynimbolinin (**95**).

(-)-O-methylcubebin (**101**), (-)-O-benzylcubebin (**102**) and (-)-O-acetylcubebin (**103**), 1 α -trigloyoxy-3 α -acetoxy-7 α -hydroxyl-12 β -ethoxynimbolinin (**104**)

CONCLUSION

P. gingivalis is one of the pathogenic bacteria found in many dental diseases. It causes other diseases such as alzheimer's, rheumatoid arthritis, diabetes, and pneumonia. The use of natural materials is one of the efforts to find natural antibacterial agents for the treatment of *P. gingivalis* infection. Several groups of compounds such as phenolics, flavonoids, terpenoids, fatty acids, and other compounds have been reported to provide specific antibacterial mechanisms against *P. gingivalis*. Several compounds from different plants have also been reported to provide excellent activity to inhibit *P. gingivalis*.

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REFERENCES

1. Bregaint S, Boyer E, Fong SB, et al. *Porphyromonas gingivalis* outside the oral cavity. *Odontology*, 2022;110: 1-19
2. Aoyama N, Kure K, Minabe M, Izumi Y. *Increased heart failure prevalence in patients with a high antibody level against periodontal pathogen*. *International Heart Journal*, 2019;60: 1142-6
3. Gasmi Benahmed A, Kumar Mujawdiya P, Noor S, Gasmi A. *Porphyromonas Gingivalis in the Development of Periodontitis: Impact on Dysbiosis and Inflammation*. *Arch Razi Inst*, 2022;77: 1539-51
4. Selbach S, Klocke A, Peters U, et al. *Microbiological and Clinical Effects of a Proanthocyanidin-enriched Extract from Rumex acetosa in Periodontally Healthy Carriers of Porphyromonas gingivalis: a Randomized Controlled Pilot Study*. *Planta Medica*, 2023;89: 1052-62
5. Afzoon S, Amiri MA, Mohebbi M, Hamedani S, Farshidfar N. *A systematic review of the impact of Porphyromonas gingivalis on foam cell formation: Implications for the role of periodontitis in atherosclerosis*. *BMC Oral Health*, 2023;23: 481
6. Pilatti F, Isolani R, Valone L, et al. *Microstructured Polymer System Containing Proanthocyanidin-Enriched Extract from Limonium brasiliense as a Prophylaxis Strategy to Prevent Recurrence of Porphyromonas gingivalis*. *Planta Medica*, 2023;89: 1074-86
7. Kang N, Zhang Y, Xue F, et al. *Periodontitis induced by Porphyromonas gingivalis drives impaired glucose metabolism in mice*. *Frontiers in Cellular and Infection Microbiology*, 2022;12: 998600
8. Mysak J, Podzimek S, Sommerova P, et al. *Porphyromonas gingivalis: major periodontopathic pathogen overview*. *Journal of immunology research*, 2014;2014:
9. Darveau R, Hajishengallis G, Curtis M. *Porphyromonas gingivalis as a potential community activist for disease*. *Journal of dental research*, 2012;91: 816-20
10. Benedyk M, Mydel PM, Delaleu N, et al. *Gingipains: critical factors in the development of aspiration pneumonia caused by Porphyromonas gingivalis*. *Journal of innate immunity*, 2016;8: 185-98
11. Olsen I, Yilmaz Ö. *Possible role of Porphyromonas gingivalis in orodigestive cancers*. *Journal of oral microbiology*, 2019;11: 1563410
12. Mei F, Xie M, Huang X, et al. *Porphyromonas gingivalis and its systemic impact: current status*. *Pathogens*, 2020;9: 944
13. Olsen I, Kell DB, Pretorius E. *Is Porphyromonas gingivalis involved in Parkinson's disease?* *European Journal of Clinical Microbiology & Infectious Diseases*, 2020;39: 2013-8
14. Carrol DH, Chassagne F, Dettweiler M, Quave CL. *Antibacterial activity of plant species used for oral health against Porphyromonas gingivalis*. *PLoS One*, 2020;15: e0239316
15. Chen D, Nie M, Fan M-w, Bian Z. *Anti-inflammatory activity of curcumin in macrophages stimulated by lipopolysaccharides from Porphyromonas gingivalis*. *Pharmacology*, 2008;82: 264-9
16. Palombo EA. *Traditional medicinal plant extracts and natural products with activity against oral bacteria: potential application in the prevention and treatment of oral diseases*. *Evidence-based complementary and Alternative Medicine*, 2011;2011:
17. Srirangarajan S, Mundargi RC, Ravindra S, et al. *Randomized, Controlled, Single-Masked, Clinical Study to Compare and Evaluate the Efficacy of Microspheres and Gel in Periodontal Pocket Therapy*. *Journal of periodontology*, 2011;82: 114-21
18. Yoshimura F, Murakami Y, Nishikawa K, Hasegawa Y, Kawaminami S. *Surface components of Porphyromonas gingivalis*. *Journal of periodontal research*, 2009;44: 1-12

19. Grenier D, Imbeault S, Plamondon P, et al. *Role of gingipains in growth of Porphyromonas gingivalis in the presence of human serum albumin*. Infection and immunity, 2001;69: 5166-72
20. Xu W, Zhou W, Wang H, Liang S. *Roles of Porphyromonas gingivalis and its virulence factors in periodontitis*. Advances in protein chemistry and structural biology, 2020;120: 45-84
21. Jia L, Han N, Du J, et al. *Pathogenesis of important virulence factors of Porphyromonas gingivalis via toll-like receptors*. Frontiers in cellular and infection microbiology, 2019;9: 262
22. Ardila C-M, Bedoya-García J-A. *Antimicrobial resistance of Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis and Tannerella forsythia in periodontitis patients*. Journal of global antimicrobial resistance, 2020;22: 215-8
23. Singhrao SK, Olsen I. *Assessing the role of Porphyromonas gingivalis in periodontitis to determine a causative relationship with Alzheimer's disease*. Journal of oral microbiology, 2019;11: 1563405
24. Kanagasingham S, Chukkapalli SS, Welbury R, Singhrao SK. *Porphyromonas gingivalis is a strong risk factor for Alzheimer's disease*. Journal of Alzheimer's disease reports, 2020;4: 501-11
25. Costa MJF, de Araújo IDT, da Rocha Alves L, et al. *Relationship of Porphyromonas gingivalis and Alzheimer's disease: A systematic review of pre-clinical studies*. Clinical oral investigations, 2021;25: 797-806
26. Dominy SS, Lynch C, Ermini F, et al. *Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors*. Science advances, 2019;5: eaau3333
27. Muñoz-Atienza E, Flak M, Sirr J, et al. *The P. gingivalis autocitrullinome is not a target for ACPA in early rheumatoid arthritis*. Journal of dental research, 2020;99: 456-62
28. Kaur S, White S, Bartold P. *Periodontal disease and rheumatoid arthritis: a systematic review*. Journal of dental research, 2013;92: 399-408
29. Gómez-Bañuelos E, Mukherjee A, Darrah E, Andrade F. *Rheumatoid arthritis-associated mechanisms of Porphyromonas gingivalis and Aggregatibacter actinomycetemcomitans*. Journal of clinical medicine, 2019;8: 1309
30. Burmester GR, Pope JE. *Novel treatment strategies in rheumatoid arthritis*. The Lancet, 2017;389: 2338-48
31. He J, Li X, Wang Z, et al. *Therapeutic anabolic and anticatabolic benefits of natural Chinese medicines for the treatment of osteoporosis*. Frontiers in pharmacology, 2019;10: 1344
32. McIntyre HD, Catalano P, Zhang C, et al. *Gestational diabetes mellitus*. Nature reviews Disease primers, 2019;5: 47
33. Ohtsu A, Takeuchi Y, Katagiri S, et al. *Influence of Porphyromonas gingivalis in gut microbiota of streptozotocin-induced diabetic mice*. Oral Diseases, 2019;25: 868-80
34. Hiroshima Y, Sakamoto E, Yoshida K, et al. *Advanced glycation end-products and Porphyromonas gingivalis lipopolysaccharide increase calprotectin expression in human gingival epithelial cells*. Journal of Cellular Biochemistry, 2018;119: 1591-603
35. Watanabe N, Yokoe S, Ogata Y, Sato S, Imai K. *Exposure to Porphyromonas gingivalis induces production of proinflammatory cytokine via TLR2 from human respiratory epithelial cells*. Journal of Clinical Medicine, 2020;9: 3433
36. Madalli R, Kheur S, Reddy MG, Kheur M, Mahalle A. *Assessment of role of Porphyromonas gingivalis as an aggravating factor for chronic obstructive pulmonary disease patients with periodontitis*. Dental Hypotheses, 2016;7: 100-6
37. Lancer F, Laribi R, Tamendjari A, et al. *Olive oils from Algeria: Phenolic compounds, antioxidant and antibacterial activities*. Grasas y aceites, 2014;65: e001-e
38. Karygianni L, Cecere M, Skaltsounis AL, et al. *High-level antimicrobial efficacy of representative Mediterranean natural plant extracts against oral microorganisms*. BioMed Research International, 2014;2014: 839019
39. Musa W, Mohd N, Zainal A. *Antibacterial Activity of Phenolic Compounds in Olive Oil Extracts on Periodontopathogenic Oral Bacteria*. Archives of Orofacial Sciences, 2022:
40. Kumbar VM, Peram MR, Kugaji MS, et al. *Effect of curcumin on growth, biofilm formation and virulence factor gene expression of Porphyromonas gingivalis*. Odontology, 2021;109: 18-28
41. Mandroli PS, Bhat K. *An in-vitro evaluation of antibacterial activity of curcumin against common endodontic bacteria*. Journal of Applied Pharmaceutical Science, 2013;3: 106-8
42. Sha AM, Garib BT. *Antibacterial effect of curcumin against clinically isolated Porphyromonas gingivalis and connective tissue reactions to curcumin gel in the subcutaneous tissue of rats*. BioMed research international, 2019;2019:
43. Izui S, Sekine S, Maeda K, et al. *Antibacterial activity of curcumin against periodontopathic bacteria*. Journal of periodontology, 2016;87: 83-90
44. Shahzad M, Millhouse E, Culshaw S, et al. *Selected dietary (poly) phenols inhibit periodontal pathogen growth and biofilm formation*. Food & function, 2015;6: 719-29

45. Wang Y, Zhang Y, Shi Y-q, et al. *Antibacterial effects of cinnamon (Cinnamomum zeylanicum) bark essential oil on Porphyromonas gingivalis*. Microbial pathogenesis, 2018;116: 26-32
46. Eltigani SA, Eltayeb MM, Ishihara A, Arima J. *Isolates from Monechma ciliatum seeds' extract hampered Porphyromonas gingivalis hemagglutinins*. Journal of food biochemistry, 2019;43: e13029
47. Park M, Bae J, Lee DS. *Antibacterial activity of [10]-gingerol and [12]-gingerol isolated from ginger rhizome against periodontal bacteria*. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 2008;22: 1446-9
48. Rivero-Cruz BE, Esturau N, Sánchez-Nieto S, et al. *Isolation of the new anacardic acid 6-[16' Z-nonadecenyl]-salicylic acid and evaluation of its antimicrobial activity against Streptococcus mutans and Porphyromonas gingivalis*. Natural product research, 2011;25: 1282-7
49. Karygianni L, Cecere M, Argyropoulou A, et al. *Compounds from Olea europaea and Pistacia lentiscus inhibit oral microbial growth*. BMC complementary and alternative medicine, 2019;19: 1-10
50. Miyake Y, Hiramitsu M. *Isolation and extraction of antimicrobial substances against oral bacteria from lemon peel*. Journal of food science and technology, 2011;48: 635-9
51. Ho KY, Tsai CC, Chen CP, Huang JS, Lin CC. *Antimicrobial activity of honokiol and magnolol isolated from Magnolia officinalis*. Phytotherapy Research, 2001;15: 139-41
52. Kugaji MS, Kumbhar VM, Peram MR, et al. *Effect of Resveratrol on biofilm formation and virulence factor gene expression of Porphyromonas gingivalis in periodontal disease*. Apmis, 2019;127: 187-95
53. Bodet C, Epifano F, Genovese S, Curini M, Grenier D. *Effects of 3-(4'-geranyloxy-3'-methoxyphenyl)-2-trans propenoic acid and its ester derivatives on biofilm formation by two oral pathogens, Porphyromonas gingivalis and Streptococcus mutans*. European journal of medicinal chemistry, 2008;43: 1612-20
54. Rodanant P, Khetkam P, Suksamrarn A, Kuvatanasuchati J. *Coumarins and flavonoid from Murraya paniculata (L.) Jack: antibacterial and anti-inflammation activity*. Pakistan journal of pharmaceutical sciences, 2015;28: 1947-51
55. Feldman M, Grenier D. *Cranberry proanthocyanidins act in synergy with licochalcone A to reduce Porphyromonas gingivalis growth and virulence properties, and to suppress cytokine secretion by macrophages*. Journal of applied microbiology, 2012;113: 438-47
56. Mahizan NA, Yang S-K, Moo C-L, et al. *Terpene derivatives as a potential agent against antimicrobial resistance (AMR) pathogens*. Molecules, 2019;24: 2631
57. Zengin H, Baysal AH. *Antibacterial and antioxidant activity of essential oil terpenes against pathogenic and spoilage-forming bacteria and cell structure-activity relationships evaluated by SEM microscopy*. Molecules, 2014;19: 17773-98
58. Chen J, Jiang Q-D, Chai Y-P, et al. *Natural terpenes as penetration enhancers for transdermal drug delivery*. Molecules, 2016;21: 1709
59. da Silva Moraes T, Leandro LF, Santiago MB, et al. *Assessment of the antibacterial, antivirulence, and action mechanism of Copaifera pubiflora oleoresin and isolated compounds against oral bacteria*. Biomedicine & Pharmacotherapy, 2020;129: 110467
60. Osawa K, Yasuda H, Morita H, Takeya K, Itokawa H. *Macrocarpals H, I, and J from the leaves of Eucalyptus globulus*. Journal of natural products, 1996;59: 823-7
61. Kubo M, Nishikawa Y, Harada K, et al. *Tetranorsesquiterpenoids and santalane-type sesquiterpenoids from Illicium lanceolatum and their antimicrobial activity against the oral pathogen Porphyromonas gingivalis*. Journal of Natural Products, 2015;78: 1466-9
62. Shirahata T, Ino C, Mizuno F, et al. *γ-Ionylidene-type sesquiterpenoids possessing antimicrobial activity against Porphyromonas gingivalis from Ph ellinus linteus and their absolute structure determination*. The Journal of Antibiotics, 2017;70: 695-8
63. Abrão F, Silva TS, Moura CL, et al. *Oleoresins and naturally occurring compounds of Copaifera genus as antibacterial and antivirulence agents against periodontal pathogens*. Scientific Reports, 2021;11: 4953
64. Moreti DLC, Leandro LF, da Silva Moraes T, et al. *Mikania glomerata Sprengel extract and its major compound ent-kaurenoic acid display activity against bacteria present in endodontic infections*. Anaerobe, 2017;47: 201-8
65. Rivero-Cruz JF, Zhu M, Kinghorn AD, Wu CD. *Antimicrobial constituents of Thompson seedless raisins (Vitis vinifera) against selected oral pathogens*. Phytochemistry letters, 2008;1: 151-4
66. Souza AB, de Souza MG, Moreira MA, et al. *Antimicrobial evaluation of diterpenes from Copaifera langsdorffii oleoresin against periodontal anaerobic bacteria*. Molecules, 2011;16: 9611-9
67. Li X-C, Cai L, Wu CD. *Antimicrobial compounds from Ceanothus americanus against oral pathogens*. Phytochemistry, 1997;46: 97-102
68. Kurnia D, Sumiarsa D, Dharsono HD, Satari MH. *Bioactive compounds isolated from Indonesian epiphytic plant of Sarang Semut and their antibacterial activity against pathogenic oral bacteria*. Natural Product Communications, 2017;12: 1934578X1701200814

69. Oh J-H, Jeong YJ, Koo HJ, et al. *Antimicrobial activities against periodontopathic bacteria of Pittosporum tobira and its active compound*. *Molecules*, 2014;19: 3607-16
70. Hernández DM, Díaz-Ruiz G, Rivero-Cruz BE, et al. *Ent-trachyloban-19-oic acid isolated from Iostephane heterophylla as a promising antibacterial agent against Streptococcus mutans biofilms*. *Fitoterapia*, 2012;83: 527-31
71. Carvalho TC, Simão MR, Ambrósio SR, et al. *Antimicrobial activity of diterpenes from Viguiera arenaria against endodontic bacteria*. *Molecules*, 2011;16: 543-51
72. Fukuyama N, Ino C, Suzuki Y, et al. *Antimicrobial sesquiterpenoids from Laurus nobilis L*. *Natural product research*, 2011;25: 1295-303
73. Zhang Q, Shi Y, Liu X-T, et al. *Minor limonoids from Melia toosendan and their antibacterial activity*. *Planta medica*, 2007;73: 1298-303
74. Desbois AP, Smith VJ. *Antibacterial free fatty acids: activities, mechanisms of action and biotechnological potential*. *Applied microbiology and biotechnology*, 2010;85: 1629-42
75. Fischer CL, Walters KS, Drake DR, et al. *Oral mucosal lipids are antibacterial against Porphyromonas gingivalis, induce ultrastructural damage, and alter bacterial lipid and protein compositions*. *International journal of oral science*, 2013;5: 130-40
76. Sweidan A, Chollet-Krugler M, Sauvager A, et al. *Antibacterial activities of natural lichen compounds against Streptococcus gordonii and Porphyromonas gingivalis*. *Fitoterapia*, 2017;121: 164-9
77. Yoshino N, Ikeda T, Nakao R. *Dual inhibitory activity of petroselinic acid enriched in fennel against Porphyromonas gingivalis*. *Frontiers in Microbiology*, 2022;13: 816047
78. Park N-H, Choi J-S, Hwang S-Y, et al. *Antimicrobial activities of stearidonic and gamma-linolenic acids from the green seaweed Enteromorpha linza against several oral pathogenic bacteria*. *Botanical studies*, 2013;54: 1-9
79. Kováč J, Slobodníková L, Trajčíková E, et al. *Therapeutic potential of flavonoids and tannins in management of oral infectious diseases—A review*. *Molecules*, 2022;28: 158
80. Soulissa AG, Lombardo B, Widyarman AS. *Antibacterial and Antibiofilm Efficacy of Pineapple Hump (Ananas comosus) on Porphyromonas gingivalis in vitro*. *Journal of Dentistry Indonesia*, 2021;28: 153-7
81. Pękal A, Pyrzynska K. *Evaluation of aluminium complexation reaction for flavonoid content assay*. *Food Analytical Methods*, 2014;7: 1776-82
82. Rivero-Cruz JF, Sánchez-Nieto S, Benítez G, et al. *Antibacterial compounds isolated from Byrsonima crassifolia*. *Revista Latinoamericana de Química*, 2009;37: 155-63
83. Fournier-Larente J, Morin M-P, Grenier D. *Green tea catechins potentiate the effect of antibiotics and modulate adherence and gene expression in Porphyromonas gingivalis*. *Archives of Oral Biology*, 2016;65: 35-43
84. Park K-M, Choo J-H, Sohn J-H, Lee S-H, Hwang J-K. *Antibacterial activity of panduratin A isolated from Kaempferia pandurata against Porphyromonas gingivalis*. *Food Science and Biotechnology*, 2005;14: 286-9
85. Villinski JR, Bergeron C, Cannistra JC, et al. *Pyrano-isoflavans from Glycyrrhiza uralensis with antibacterial activity against Streptococcus mutans and Porphyromonas gingivalis*. *Journal of natural products*, 2014;77: 521-6
86. Cai L, Wu CD. *Compounds from Syzygium aromaticum possessing growth inhibitory activity against oral pathogens*. *Journal of natural products*, 1996;59: 987-90
87. Iwaki K, Koya-Miyata S, Kohno K, Ushio S, Fukuda S. *Antimicrobial activity of Polygonum tinctorium Lour: extract against oral pathogenic bacteria*. *Journal of Natural Medicines*, 2006;60: 121-5
88. Ho K, Tsai C, Huang J, et al. *Antimicrobial activity of tannin components from Vaccinium vitis-idaea L*. *Journal of Pharmacy and Pharmacology*, 2001;53: 187-91
89. Yamamoto H, Ogawa T. *Antimicrobial activity of perilla seed polyphenols against oral pathogenic bacteria*. *Bioscience, biotechnology, and biochemistry*, 2002;66: 921-4
90. Löhr G, Beikler T, Podbielski A, et al. *Polyphenols from Myrothamnus flabellifolia Welw. inhibit in vitro adhesion of Porphyromonas gingivalis and exert anti-inflammatory cytoprotective effects in KB cells*. *Journal of clinical periodontology*, 2011;38: 457-69
91. Minami M, Takase H, Nakamura M, Makino T. *Effect of Lonicera caerulea var. emphylocalyx Fruit on Biofilm Formed by Porphyromonas gingivalis*. *BioMed Research International*, 2019;2019:
92. Milho C, Silva J, Guimarães R, et al. *Antimicrobials from medicinal plants: an emergent strategy to control oral biofilms*. *Applied Sciences*, 2021;11: 4020
93. Kulik EM, Thurnheer T, Karygianni L, et al. *Antibiotic susceptibility patterns of aggregatibacter actinomycetemcomitans and porphyromonas gingivalis strains from different decades*. *Antibiotics*, 2019;8: 253
94. Eltigani SA, Eltayeb MM, Bito T, et al. *Argeloside I inhibits the pathogenicity of Porphyromonas gingivalis TDC60*. *Journal of bioscience and bioengineering*, 2020;130: 644-9

95. Hioki Y, Onwona-Agyeman S, Kakumu Y, et al. *Garcinoic acids and a benzophenone derivative from the seeds of Garcinia kola and their antibacterial activities against oral bacterial pathogenic organisms*. Journal of natural products, 2020;83: 2087-92
96. Rezende K, Lucarini R, Símaro GV, et al. *Antibacterial activity of (-)-cubebin isolated from Piper cubeba and its semisynthetic derivatives against microorganisms that cause endodontic infections*. Revista Brasileira de Farmacognosia, 2016;26: 296-303
97. Zhang Q, Zheng Q-H, Liang J-Y, Qing-Shan L, Zhi-Da M. *Two new limonoids isolated from the fruits of Melia toosendan*. Chinese journal of natural medicines, 2016;14: 692-6